

The use of ozone in whole room disinfection

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The use of ozone in whole room disinfection

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Summary

Campden BRI, in collaboration with various partners (system manufacturers and users), has been studying the possible use of whole room disinfection to control pathogens and potential spoilage organisms in the food production environment (factory or process hall) for over 20 years. In this time there has been an increase in the demanded and expectation of higher standards in the control of microorganisms within the food production environment. This, coupled with the identification of environmentally persistent strains of pathogens (Holah *et al* 2002, Holah *et al* 2004) has led to a significant interest in the use of whole room disinfection techniques to supplement routine cleaning and disinfection.

A range of whole room decontamination systems are available commercially; this report is a summary update on the use of ozone in whole room disinfection and the development of supportive data for its use within the food and drink industry. This report summaries work undertaken to investigate the efficacy of ozone for whole room disinfection against microorganisms attached to surfaces, both in the laboratory and in the factory environment.

The microorganisms assessed in laboratory trials at Campden BRI were *Listeria monocytogenes*, *Pseudomonas aeruginosa* and *Staphylococcus aureus*; they were selected because of their potential presence, and in some cases persistence, in the food factory environment. This work was also undertaken to examine the effect of different spatial orientations of contaminated surfaces upon the efficacy of whole room disinfection techniques.

A relationship between ozone concentration, log reduction of microorganisms and contact time was established, though the effect of ozone on each of the three vegetative strains tested varied. There was a marked difference in organism sensitivity with, at 20ppm, *S. aureus* being most resistant, followed by *P. aeruginosa* and then *L. monocytogenes* being most sensitive. A reduction of >4 logs was achieved for *L. monocytogenes* at an ozone concentration of 20ppm within a contact or dwell time of 1h (excluding ozone concentration build-up and breakdown times which combined together took 3 h process time) (Middleton 2010).

There was little practical difference in log reductions achieved between orientations (horizontal, vertical and underneath surfaces) (Malinowska and Holah 2007, Middleton 2010) or, as shown in later studies, between surfaces within tubes of a test rig (Plate 1). As such, ozone can effectively penetrate every part of a room, including sites that might prove difficult to gain access to with conventional liquids and manual disinfection procedures.

The major disadvantage of using gases, such as ozone, is the potential toxicity to workers at high concentrations or extended exposures, which precludes using them in areas where people are working. The techniques can therefore only be used in areas that can be isolated, or sealed off during the decontamination process, and well ventilated after, or where the gas can be allowed enough time to break down naturally or using a catalyst.

Field trials in which ozone was used as a room decontaminant showed little effect over 1 and 2 days using an ozone concentration of 5 -8ppm for contact times of 40 to 60 minutes (Malinowska and Holah, 2007, Middleton 2010) and the overall reduction in counts after disinfection were probably less than 1 log order. This may be because, for all areas in which ozone is used, oxidisable material within the room may create an ozone demand which must be satisfied by oxidation before any significant decontamination of microorganisms can occur.

However, the results for a pizza manufacturer (3.2.1) in which ozone was used at 8ppm for 40 minutes over three days showed a downward trend in the numbers of microorganisms present, both before cleaning and after cleaning and disinfection.

In a 4 week field trial at a sandwich manufacturer, (3.2.2), TVC counts after disinfection for ozone treated food contact surfaces compared favourably to the post disinfection counts of combined disinfectant approval trials conducted at other chilled food plants.

A longer 15-week validation trial in 2 halls of a dough manufacturing site (3.2.3) provided supporting evidence of the potential benefit of a downward trend in production environment contamination post cleaning.

Importantly, no adverse effects were reported with ozone at appropriate concentrations and contact times on the structure and fabric of the building or equipment following installation of ozonation equipment for daily treatment of 8ppm for 30 minutes (sandwich factory) or weekly treatment of 6ppm for 3 hours with natural decay (dough factory).

The results of laboratory and field trials indicate that ozone at appropriate concentrations and contact times has the potential to be an effective environmental disinfectant. Any field trials of these systems must be safely undertaken and should be of sufficient length (time / number of applications) to effectively evaluate decontamination performance.

Acknowledgement

Campden BRI would like to thank Dow Microbial Control for sponsoring this report update, and all those factories who submitted field trial and validation data for inclusion in this review.

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1. BACKGROUND

Food can be exposed to microbiological cross-contamination from food contact surfaces via direct contact, and from non food contact surfaces via vectors such as the air, people etc, which may give rise to food spoilage and safety issues. The traditional approach to controlling such contamination has been to target specific sites within the manufacturing environment with cleaning and disinfection regimes. The primary focus is typically on food production equipment and drains. The remaining food production environment/processing area, whilst cleaned, may not be routinely disinfected. This targeted approach may have been sufficient to maintain day-to-day control of contamination, but does not eliminate all of the organisms within a production environment and, in some instances, microbial strains have become persistent in food factories, surviving for several years (Holah *et al* 2002, Holah *et al* 2004). Clearly, these organisms present a cross contamination risk and if there was any loss of hygiene control in these factories, these organisms could present a risk to product safety.

In high risk food processing areas, thorough disinfection of surfaces is required in order to reduce the numbers of microorganisms and to prevent transmission of these contaminants. Through the regular use of various disinfection techniques it is believed that the “whole room” can be decontaminated. This will reduce the number of environmental microorganisms in the production areas (bioburden), and may also help reduce the incidence of persistent strains; thus reducing the risk that these organisms would contaminate product and improving the quality and safety of the food being produced, thereby reducing wastage and increasing profitability.

One of the systems investigated at Campden BRI and successfully used in factories is the application of ozone gas in a non-condensing humid environment (Holah *et al* 2002, Holah *et al* 2004, Middleton 2010).

2. INTRODUCTION

There is now a growing desire to supplement traditional, targeted chemical disinfection with alternative approaches which will control micro-organisms in the greater food processing environment, be it a wet, high care or dry production environment. This technique is termed “whole room disinfection”. Novel disinfection techniques that are able to disinfect whole areas have been implemented in the pharmaceutical, clinical and now food sectors; one of those techniques is the use of ozone gas within a non condensing humidified atmosphere.

2.1 Ozone: what is it?

Ozone is the triatomic form of oxygen and is found in our atmosphere (the majority of this is in the “ozone layer”). It is unstable, naturally breaking down into molecular oxygen. The rate of break down is dependent upon environmental conditions such as temperature, humidity, and pollution but its approximate half life is 20 minutes in air and 30 minutes in water.

Ozone is a strong oxidiser and highly reactive. This, combined with penetrability and spontaneous decomposition into a non-toxic product, make ozone a viable disinfectant for use in food production areas.

2.2 Health and safety

Ozone is a toxic gas and worker exposure should be controlled. Table 1 gives examples of worker exposure limits for ozone in the US and UK. Many people can detect ozone in the air via smell at around 0.03 ppm, far below the recommended exposure limits, and at higher concentrations it can lead to headaches along with irritation to the eyes and respiratory tract.

Whilst ozone is a toxic gas, it is considered environmentally friendly as it readily breaks down to molecular oxygen (half life of approx. 20 minutes) and leaves no chemical residues.

Table 1: Ozone exposure (limits in US and UK)

USA	Occupational Safety and Health Administration (OSHA)	PEL is 0.1 ppm	OSHA 29 CFR 1910.1000(a)(2) Table Z-1
	Occupational Safety and Health Administration (OSHA)	STEL 0.3ppm	OSHA PEL Project Documentation 1988 TABLE AC-1 PERMISSIBLE EXPOSURE LIMITS FOR CHEMICAL CONTAMINANTS
	National Institute for Occupational Safety and Health (NIOSH)	Immediately Dangerous to Life or Health Concentration (IDLH) = Ozone 5 ppm Recommended STEL = 0.1 ppm ceiling	NTIS Pub PB-94-195047 (1995)
	American Conference of Governmental Industrial Hygienists	Heavy work: 0.05 ppm 8-hour TWA Moderate work: 0.08 ppm 8-hour TWA Light work: 0.1 ppm 8-hour TWA All workloads: 0.2 ppm 2-hour TWA	American Conference of Governmental Industrial Hygienists (ACGIH) Threshold Limit Value (TLV) (2001)
UK	HSE	STEL 0.2ppm LTEL (none given)	EH40/2005 Workplace exposure limits document
	UK National Air Quality Standards website*	air quality objective of 50ppb (0.05ppm) as the 8-hour mean	: www.airquality.co.uk/archive/standards.php#band

Legend

- *PEL Permissible Exposure Limit. An employee's exposure to any substance in OSHA Table z-1 shall not exceed the 8-hour Time Weighted Average given for that substance in any 8 hour work shift for a 40-hour work week.*
- *STEL Short Term Exposure Limit. An employee's exposure shall not exceed this Time Weighted Average over 15 minutes.*
- *TWA Time Weighted Average*
- *LTEL Long Term Exposure Limit*
- *ppm Parts per million*
- *ppb Parts per billion*
- ** This is an objective for "natural" ozone in the air and should not be used as an LTEL*

2.3 How is ozone made and applied?

Due to its reactive, unstable nature, ozone is produced at the point of use. Ozone generators effectively pass air or oxygen through a high-energy source within the equipment and the resulting physicochemical reaction leads to the formation of ozone that can be used for area or surface decontamination. Widely used high-energy sources include UV light (produce $\leq 0.5\%$ ozone used in spas and swimming pool water) and electrochemical cells or corona discharge ozone generation (3-6% ozone). A corona is formed by an electrical discharge around a gas (often air but oxygen can also be used), which causes ionisation of the gas and consequently the formation of ozone. The production of ozone is most effective in a temperature-controlled environment, since the stability of ozone decreases as the temperature increases.

Its use typically involves humidification (70%-90% RH) of the environment followed by the application of ozone in the humidified environment, which is maintained for the required contact time, and finally reduction of ozone/humidification to normal levels, via either air replacement, natural breakdown or use of a catalyst system to actively break down ozone.

2.4 Anti microbial uses and microbial susceptibility

Surfaces can be treated using ozone dissolved in water, or as a gas in a humid atmosphere. Microorganisms inherently vary in their sensitivity to ozone, with factors such as temperature, humidity, the presence of chemicals, and the amount of organic matter surrounding the cell greatly affecting the degree of inactivation. At the concentrations typically used, ozone is an effective bactericide and virucide (Maillard *et al.* 2012, Hudson *et al.* 2007) whilst mycobacteria and bacterial spores have been shown to be less susceptible. Effective sporicidal activity is only seen at high relative humidity (75 to 95%) and high concentrations with long contact times (Dusseau *et al.* 2012). Yeasts and moulds have been reported to have a wide range of resistance profiles; however, ozone has been demonstrated to control post harvest spoilage of cereals, grains and fruit as well as reducing mould and yeast contamination of cheese during ripening (Boisrobert 2002, Siqueria and Botelho da Silva, 2008). However, mould spores are reportedly less resistant than bacterial spores to ozone and disinfectants generally.

A review of papers assessing the use of ozone applied as a gas in humid atmospheres for different applications is shown below (Table 2).

Table 2
Summary of literature search results looking at ozone applied via gas for surface disinfection in food industry (2000 – 2013)

Title	Author	Source	Notes	Year
Bactericidal properties of ozone and its potential application as a terminal disinfectant	Moore, G., Griffiths, C. and Peters, A.	Journal of Food Protection 63(8),1100-1106	2ppm ozone, 20°C, 77%RH produced 2 - 7 log reduction against a range of bacteria in the presence and absence of UHT milk	2000
The evaluation of ozone on airborne and surface disinfection	Taylor, J. and Chana, D.	R&D report No 109 Campden BRI	Demonstrated efficacy against <i>P. aeruginosa</i> in the air and on stainless steel surfaces	2000
Inactivation kinetics of foodborne spoilage and pathogenic bacteria by ozone.	Kim, J.-G. & Yousef, A.E.	Journal of Food Science, 65(3), 521-528.	Ozone was tested against <i>P. fluorescens</i> , <i>E. coli</i> O157:H7, <i>L. mesenteroides</i> and <i>L. monocytogenes</i> . Survivor plots in the continuous system were linear initially, followed by a concave downward pattern. Exposure of bacteria to ozone at 2.5 ppm for 40 s caused a 5 to 6 log decrease in count. Resistance of tested bacteria to ozone followed this descending order: <i>E. coli</i> O157:H7, <i>P. fluorescens</i> , <i>L. mesenteroides</i> , and <i>L. monocytogenes</i> .	2000
US regulatory review of the use of ozone in the food industry	Biosrobert, C.	Agricultural and food processing applications of ozone as an antimicrobial	Reviews use of ozone to control spoilage organisms including fungi	2002
Gaseous ozone treatment inactivates <i>Listeria innocua</i> <i>in vitro</i> .	Fan, L., Song, J., McRae, K.B., Walker, B.A. and Sharpe, D	Journal of Applied Microbiology, 103, 2657-2663.	Average time for a 2 log reduction of <i>Listeria innocua</i> on solid media was 1.3 hours at 20°C, and 2.5 hours at 5°C	2007
Whole room disinfection - potential for environmental pathogen control	Malinowska, A. and Holah, JT.	New Food (no5) 2007 22-26	Ozone efficacy against <i>S. aureus</i> , <i>P. aeruginosa</i> , <i>L. monocytogenes</i>	2007
Use of ozone in food industries for reducing the environmental impact of cleaning and disinfection activities	Pascaul, A., Llorca, I. and Canut, A.	Trends in Food Science & Technology 18 (Suppl. 1), 2007 s29-s35	Ozone use in meat processing plant and wineries	2007

Title	Author	Source	Notes	Year
Use of ozone in industrial cold rooms to control yeast and moulds during parmesan cheese ripening	Siqueria Lanita, C. de Botelho and da Silva, S.	Brazilian Journal of Food Technology; 11(3), 182-189	0.03mg/L ozone was shown to reduce air contamination and surface spoilage of product	2008
Reduction by gaseous ozone of <i>Salmonella</i> and microbial flora associated with fresh-cut cantaloupe	Selma, M.V., Ibanez, A.M., Cantwell, M. and Suslow, T.	Food Microbiology; 25(4) 558-565	Gaseous ozone is an effective option in risk reduction and spoilage control of fresh and fresh cut melon. Ozone treatment combined with rapid drying reduces persistence of <i>Salmonella</i> on surface and reduces risk of transference from rind to flesh during cutting	2008
The case for ozone	Brandit, J.	Food Quality (Dec/Jan) 2009	Review	2009
Whole room disinfection	Middleton, K.	Food and Beverage International; Vol 8(6), 2009, 46-47	Efficacy of whole room disinfection methods including ozone	2009
Whole room disinfection	Middleton, K.	Campden BRI R&D report 299	Efficacy of whole room disinfection methods including ozone	2010
Application of gaseous ozone to inactivate <i>Bacillus cereus</i> in processed rice.	Shah N.N.A.K. Rahman, R.A. and Chuan, L.T.	Journal of Food Process Engineering 34(6), 2220-2232	Approximately 2 log reduction at 0.4ppm 20C 50%RH 420 min. 1.63 log reduction 0.3ppm 20C 50% RH 420 min	2011
Inactivation of <i>Listeria</i> , <i>Salmonella</i> Typhimurium and <i>Escherichia coli</i> O157H7 on surface and stem scar areas of tomatoes using in package ozonation	Xuetong Fan, Sokoral, K. J. B., Engermann, J, Gutler, J.B. and Yanhong Liyu	Journal of food protection; 75(9), 1611-1618	Bacteria responded differently to ozonation <i>Listeria</i> susceptible ≥ 4 log reduction within 40s <i>E. coli</i> and <i>Salmonella</i> 2-3 log reduction after 2-3 min (1000ppm <i>in-situ</i> after 1 minute)	2012
Mould control by ozonation in ripening cheese room	Troller Pinto, A., Scmidit, V. and Aparecida Raimundo. S.	Acta Scientiae Veterinariae, 35 (3), 333-337	Control of environmental and surface fungi 0.74 log on cheese surface, 0.91 log reduction on shelf surface and 1.5 log reduction in air	2013
Disinfection of selected vegetables under non-thermal treatments: chlorine, citric acid, ultraviolet light and ozone	Bermudez-Aguirre, D. and Barbosa-Canovas, G.V.	Food Control 29 (1), 82-90	5 ppm ozone demonstrated <i>E. coli</i> log reductions of 2.2 log on tomatoes, but affected greenness of lettuce	2013

It can be seen from the data in Table 2 that the efficacy of ozone like other disinfectants is dependent upon a range of factors: concentration, contact time, temperature, presence of interfering organic matter, and the target organisms.

However, all references demonstrate an added benefit when used in an appropriate manner.

In a number of tests *L. monocytogenes* seems to be the most susceptible organism tested

2.5 General consideration of use

The critical factors to address before using techniques such as whole room disinfection via gaseous or aerosolised disinfectants include:

- identifying areas where the decontamination processes can be applied
- health and safety issues related to using the technique, e.g. staff exposure
- effects on the fabric of the equipment and the building, e.g. potential corrosion.

These can be controlled through risk assessments and the implementation of management procedures to monitor ozone concentrations and dispersal (both of the areas treated and adjacent areas), control of access to treated areas, and the monitoring of efficacy.

The techniques can be used daily, weekly, or monthly, or on an ad-hoc basis as a reaction to a particular issue. The frequency of application, concentrations of ozone applied, contact time and target organism along with the type of environmental contamination to be encountered have all been shown to affect efficacy. Therefore, it is imperative that the method is validated in some way to be sure that it is effective.

3. SUPPORTING DATA

Various laboratory and field trials are quoted above in Table 2, which also includes laboratory and field trials carried out by Campden BRI.

3.1 Laboratory trials at Campden BRI

Laboratory trials (Malinowska and Holah 2007, and Middleton, 2010) looked at *Pseudomonas aeruginosa*, *Listeria monocytogenes*, and *Staphylococcus aureus*. The purpose of these laboratory trials was to further develop the method to examine whether the systems under assessment were able to decontaminate surfaces, irrespective of orientation, throughout the whole room, providing similar levels of log reduction, and to determine ozone efficacy. The laboratory trials protocol was based on the European Norm surface disinfectant test method BS EN 13697: 2001 - Chemical disinfectants and antiseptics - Quantitative non-porous surface test for evaluation of the bactericidal activity and/or

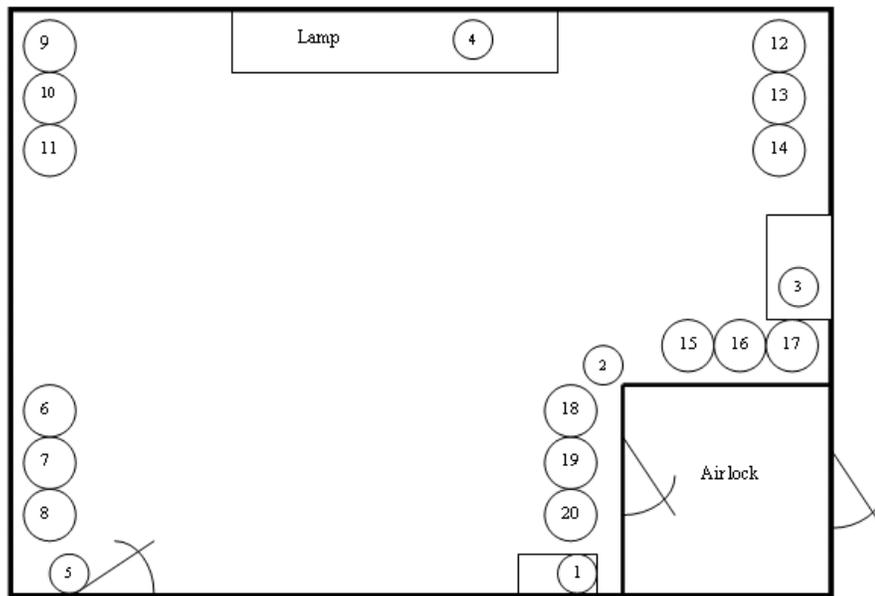
fungicidal activity of chemical disinfectants used in food, industrial, domestic and institutional areas (Anon 2001). In these experiments 0.05mL bacterial suspension (of approximately $1.0E+07$ CFU/mL) was dried onto stainless steels surfaces (2cm diameter 1.4301(EN 10088-1): stainless steel discs with grade 2B finish on both sides (in accordance with EN10088-2, gauge 1.2 mm – 1.5mm).

The discs were placed in locations shown in Figure 1.

The ozone generator was placed in the centre of the room with the dispensing head approximately 1.2m from the floor. The ozone generator was activated to implement the required decontamination process.

Figure 1 - The arrangement of test surfaces in the aerobiology laboratory

- Key:**
- | | | | |
|------------------------------------|-------------------------------------|-------------------------------------|-------------------------------------|
| - 1 = underneath: plug | - 6 = top: horizontal | - 11 = bottom: underneath | - 16 = middle: vertical facing wall |
| - 2 = floor | - 7 = middle: vertical facing wall | - 12 = top: horizontal | - 17 = bottom: underneath |
| - 3 = vertical: inside HEPA alcove | - 8 = bottom: underneath | - 13 = middle: vertical facing wall | - 18 = top: horizontal |
| - 4 = vertical: underneath lamp | - 9 = top: horizontal | - 14 = bottom: underneath | - 19 = middle: vertical facing room |
| - 5 = horizontal: above door | - 10 = middle: vertical facing room | - 15 = top: horizontal | - 20 = bottom: underneath |



During experiments, all ventilation systems in the laboratory were switched off and the room was effectively sealed to outside air movements. Any internal air currents during the trials were created by the operation of the technology under test.

The treated discs and untreated control discs were recovered into 10mL neutraliser broth with glass beads and the surviving CFU/mL were determined.

Log₁₀ reduction was determined via the formula:

$$\text{Log}_{10} \text{ reduction} = \text{mean Log}_{10} \text{ surviving CFU/disc controls} - \text{Log}_{10} \text{ surviving CFU/disc treated}$$

The log reductions achieved in all ozone trials are summarised in Figure 2.

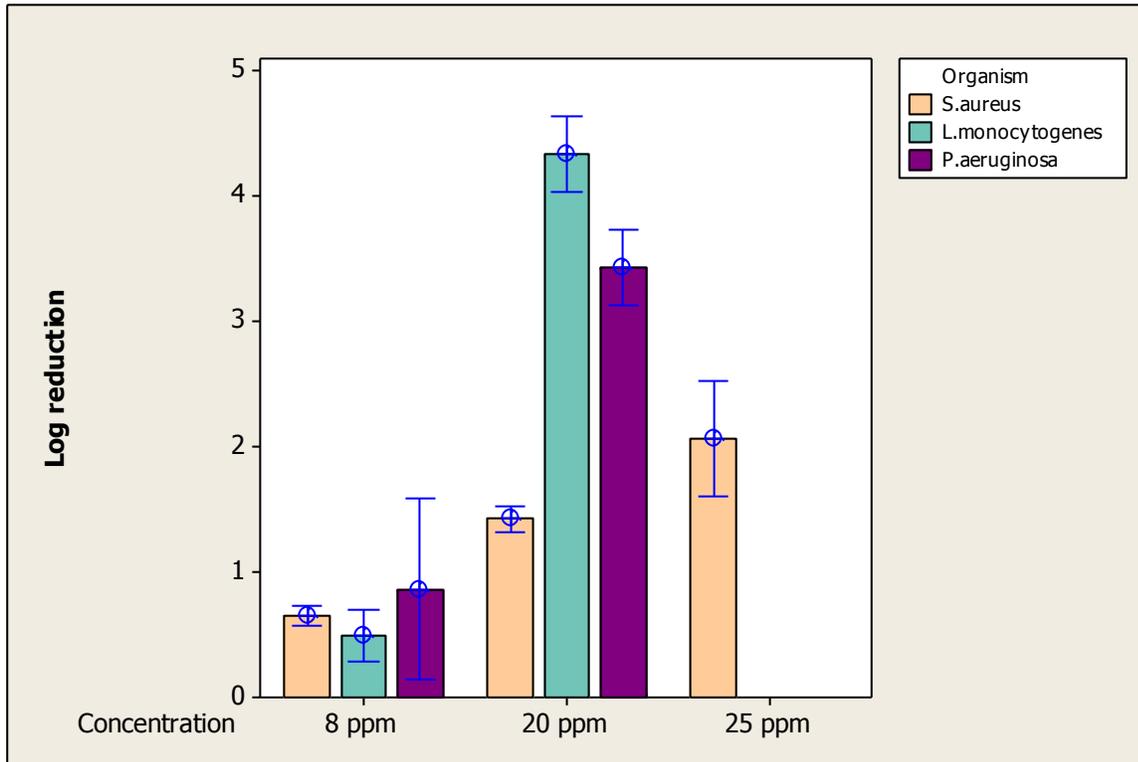


Figure 2 - Effect of gaseous ozone on microbiological mean log reduction at different concentrations from all locations/orientations (dwell time 1h)

The results in Figure 2 show two trends. Firstly, there is a clear relationship between ozone concentration and log reduction, with the log reduction for *S. aureus* ranging from 0.7 logs at 8ppm, to 1.5 logs at 20ppm and 2.1 logs at 25ppm. Secondly, the effect of ozone on the three vegetative strains tested is markedly different with, at 20ppm, *S. aureus* being most resistant, followed by *P. aeruginosa*, and *L. monocytogenes* being most sensitive. It should be noted that *L. monocytogenes* and *P. aeruginosa* were not included in the 25ppm trial.

The effect of sample orientation on log reduction at an ozone concentration of 20ppm is shown in Figure 3.

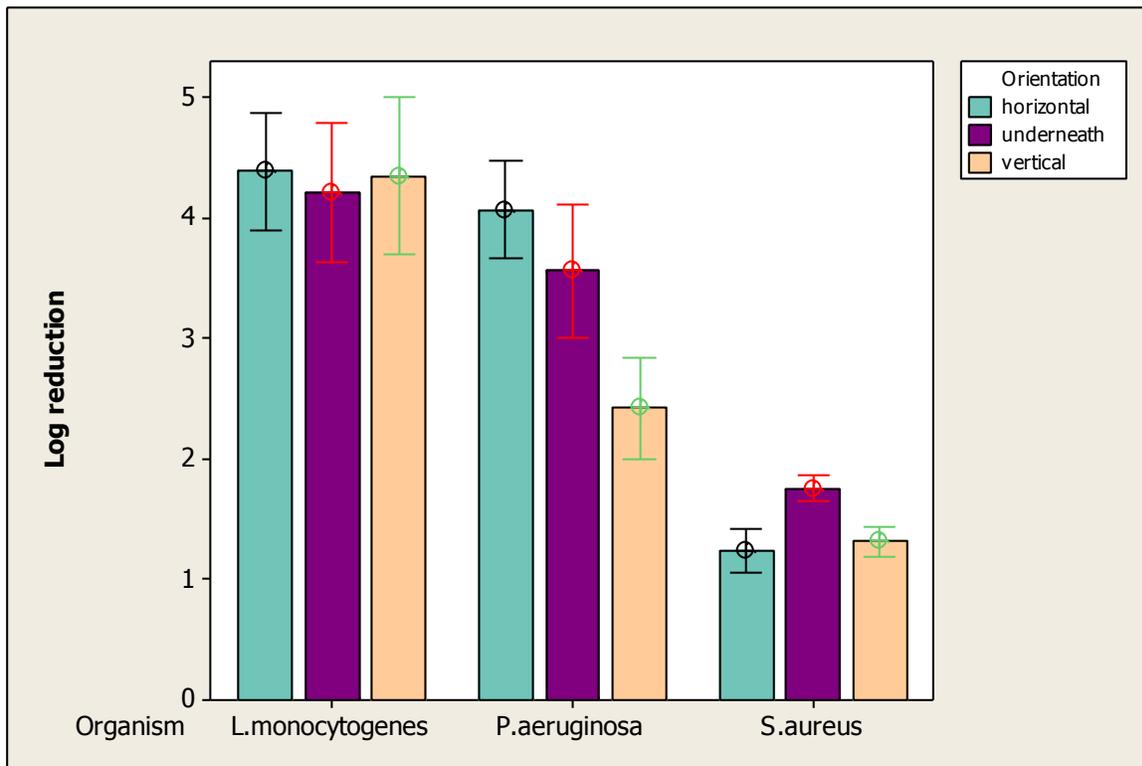


Figure 3 - Effect of gaseous O₃ on microbiological log reduction depending on orientation at 20 ppm long cycle (dwell time 1h)

There is no statistical difference in log reduction with orientation for *L. monocytogenes* ($P=0.896$); although the orientation log reductions were statistically different for both *P. aeruginosa* and *S. aureus* it was deemed that there was no practical difference (< 1 log). There is little evidence to suggest that ozone is not able to penetrate to all surfaces, irrespective of their orientation.

Further studies (2013) have demonstrated the ability for ozone to penetrate hard-to-reach spaces such as W Tubes (Plate 1, Table 3). The inoculated surfaces were placed in the red capped areas; to be effective the gas must migrate through the tubes.

Plate 1 W tubes used in aerobiology tests post 2011



Table 3: Results of laboratory trial carried out in 2013

20 ppm ozone, dwell time 30 minutes (total 55 minutes), extraction via catalyst removal	
Log mean control surface	6.76
Mean log reduction	
Mean log reduction horizontal surfaces (high) (non line of sight)	0.43
Mean count for vertical, facing room surfaces (line of sight)	0.70
Mean log count for vertical, facing wall (non line of sight)	0.63
Mean log reduction underneath surfaces (non line of sight)	0.70
Mean log reduction horizontal surfaces table top (line of sight - below dispensing head)	0.55
Mean log reductions W-Tube (non line of sight)	0.48
Mean log reduction achieved in test	0.58

Line of sight – a direct line could be drawn from the ozone dispensing head to the inoculated surface

Reduced contact time (30 minutes) was shown to produce a 0.58 mean log reduction

Variation in log reduction achieved between positions of discs and mean log reduction was <0.3 log with those discs within the W tubes

3.2 Field trials

Field trials of 1-3 treatment applications were conducted by Campden BRI in conjunction with a manufacturer of ozone in representative factories of the RTE, poultry and dry food industry (Malinowska and Holah 2007). The results from these trials were varied. In addition one longer term 4 week trial in a sandwich factory was done.

In these field trials the methodology was based on procedures developed by Campden BRI (TES-FH -013-PART2).

An independent factory validation trial (not undertaken by Campden BRI) was also carried out in two dough factory sites. This data was kindly provided by the factory involved for publication by Campden BRI.

3.2.1 Factory: Pizza manufacturer (Malinowska and Holah, 2007)

Test site: High care cook-house, room size 75m³.

Trial protocol: the trial was conducted at the end of production on three occasions. The environment was cleaned with detergent and rinsed and then terminal disinfectant treatment was replaced by ozone treatment.

Ozone treatment: 8ppm for 40 minutes + quench.

A total of 15 swabs and 15 contact plates (Table 3.2.1) were taken as detailed below. The sites were sampled by Campden BRI staff, who took samples before cleaning, after cleaning and after disinfection.

Table 3.2.1 - Swab and contact plate sites

Site Code	Sample site - product contact surfaces
1	Vegetable Tumbler (inside)
2	Shelf underneath Vegetable tumbler
3	Red switch on/off (Vegetable tumbler)
4	mixer – blade
5	mixer - bottom (inside)
6	mixer – outlet
7	Stainless steel table-top
8	mixer – panel
9	Stainless steel table – top
10	Blast Chill Room 18B door handle
Site Code	Sample site - Environmental samples
1E	Floor (stainless steel)
2E	Wall (behind mixer)
3E	Floor (red)
4E	Around drain seal to the floor
5E	Drain inside (drain's basket)

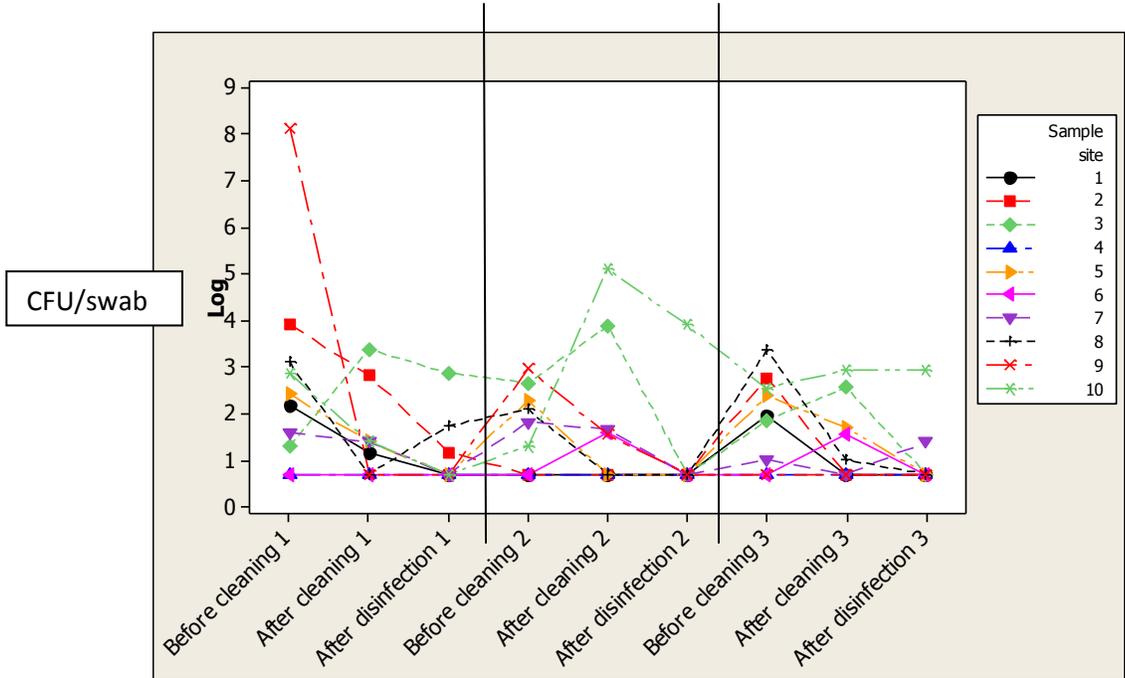


Figure 3.2.1a - Effect of gaseous O₃ treatment on microbiological log reduction of food contact surfaces within a pizza manufacture facility (Log CFU/swab recovered)

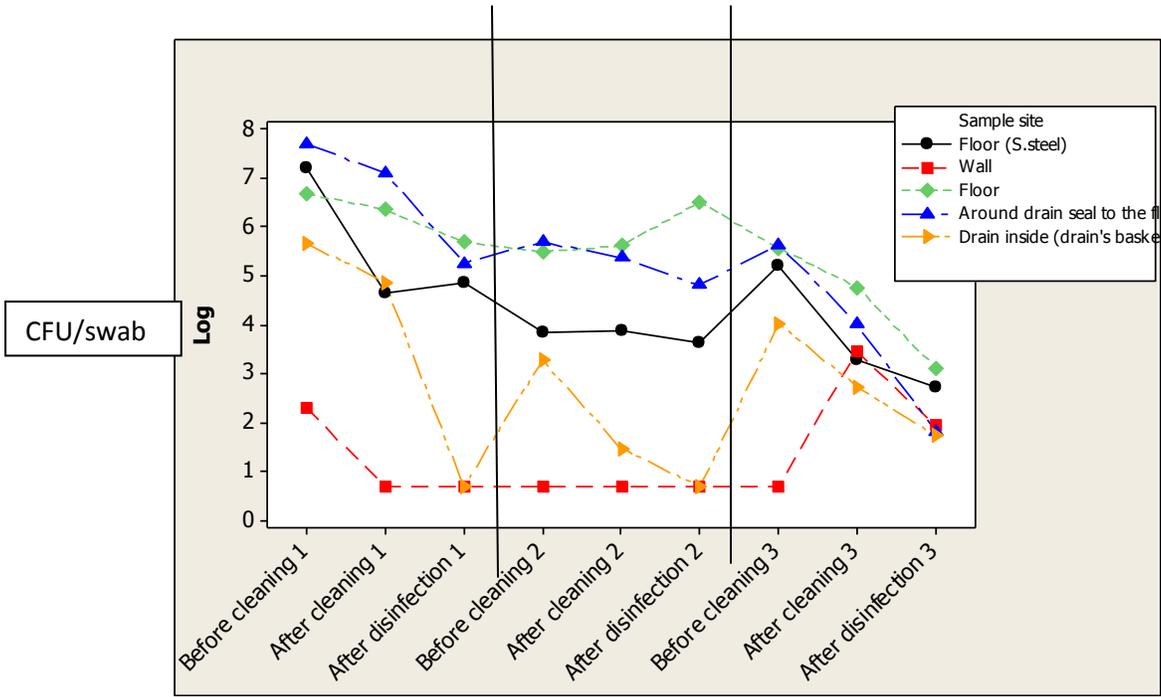


Figure 3.2.1b - Effect of gaseous O₃ treatment on microbiological log reduction of environmental surfaces within a pizza manufacture facility (log CFU/swab recovered)

The results for the food environment of the pizza factory over the three day trial (Figure 3.2.1b) show that, on each individual day, there is a downward trend in number of organisms recovered, after cleaning and after disinfection. The results of the environmental samples also indicate a downward trend for the numbers of microorganisms present before cleaning over the three day period.

Previous studies using single applications or two applications over two days have demonstrated very little effect (Middleton, 2010, Malinowska and Holah, 2007). It was speculated that the environment, even after cleaning, may have an “oxidation demand” that required meeting before the full effects on organisms and food contact surfaces, and in the manufacturing environment, were consistent. As such it is recommended that field trials to validate the efficacy of such systems be carried out over a period of multiple applications (at least 3).

3.2.2 Factory: Sandwich manufacturer (Malinowska and Holah, 2007)

Test site: High care area, room size 1080m³.

Trial protocol: routine end of production cleaning and disinfection. Gross solids removal, rinse with 3ppm ozonated water, chemical application, rinse with 3ppm ozonated water, seal room, disinfect with 8ppm for 30 minutes and quench.

The sample sites were 10 food contact surfaces of various materials of construction and 10 environmental surfaces (including walls, floors and drain areas).

Table 3.2.2 – A schematic comparison of chemical disinfection and O₃ (Malinowska and Holah, 2007)

	Pre cleaning Log CFU/swab	Post cleaning Log CFU/swab	After disinfection Log CFU/swab
Mean of counts per swab from 10 separate, 8 week duration chemical disinfectant trials (food contact surfaces) undertaken at other RTE ready meal factories.	4.73	2.80	1.30
Mean of counts per swab from 4 week duration trials using ozone as a disinfectant (food contact surfaces).	2.32	1.98	1.29
Average of counts per swab from 4 week duration trials using ozone as a disinfectant (environmental samples).	3.65	2.83	2.27

The mean counts from the four week trial on food contact surfaces were compared to the averaged data from 10 field trials using chemical disinfectants gathered over a number of years.

The results in Table 3.2.2 show that the TVC count decreased after cleaning and again after disinfection on food contact surfaces when using traditional chemical disinfectants and ozone.

The results in Table 3.2.2 show that for food contact surfaces, the mean counts after disinfection with the ozone treatment described compare favourably to the post disinfection counts of combined chemical disinfectant trials. Therefore ozone was considered to maintain control of the microflora of the food contact surfaces.

Mean TVC counts from environmental surfaces were reduced after cleaning and again after disinfection with ozone. Though they were higher than on food contact surfaces, the disinfection of the production environment is considered to reduce the risk of cross contamination and reduce the risk from persistent (resident) strains.

During the 4 week trial no adverse effects were observed on the structure and fabric of the building. The management of the factory have also reported no adverse effects since installation of the ozonation equipment (Malinowska and Holah, 2007).

3.2.3 Factory: Validation of an installed system in two dough manufacturing halls (2013)

Test site: Two manufacturing halls for two types of dough.

Trial protocol: the trial was conducted at the end of production. The environment was a dry food manufacturing one (normal relative humidity of 35%) and was appropriately cleaned prior to ozone treatment application once per week. The site considered that it had a number of persistent strains of *Listeria* spp. For the year prior to and during the trial there was no change in production volumes, methods of production or sanitisation (excluding the use of ozone) or number of staff in the areas.

Ozone treatment was 6ppm at 80% relative humidity for 4 hours with natural ozone break down (generation time 25 minutes and 4.5 hours typical natural breakdown), with a total treatment time of approximately 9.25 hours

For both halls ozone treatment was used weekly from week 1 through to week 16.

There followed a break in production between weeks 17–25 and then production was restarted in week 26. However, weekly ozone treatment was not restarted until week 35.

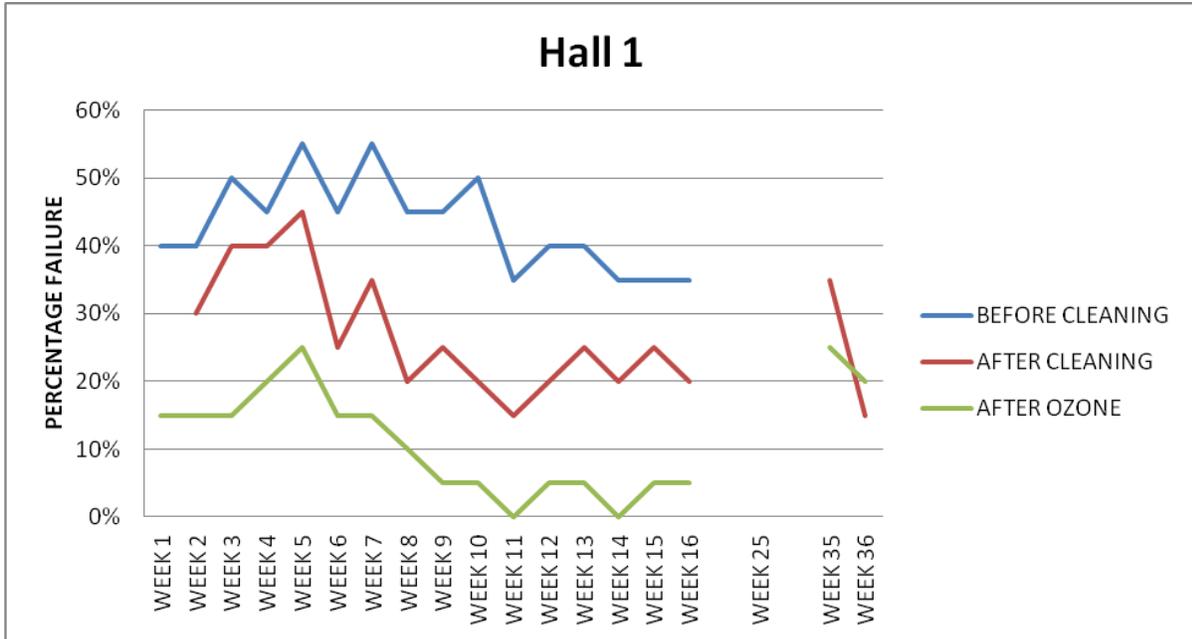
A total of 20 sites per production area (Tables 3.2.3a & b) were tested for *Listeria* spp using “3M™ Petrifilm™ Environmental Listeria Plate”. The percentage positive sites found were recorded over a 15 week period and are graphically represented below (Graphs 3.2.3a and 3.2.3b).

The results over the first 16 weeks of both trials demonstrated an overall downward trend in pre and post clean positives. The post ozonation reductions demonstrated that ozone is effective in reducing *Listeria* spp in the environment compared with post cleaning, therefore reducing the risk of cross contamination.

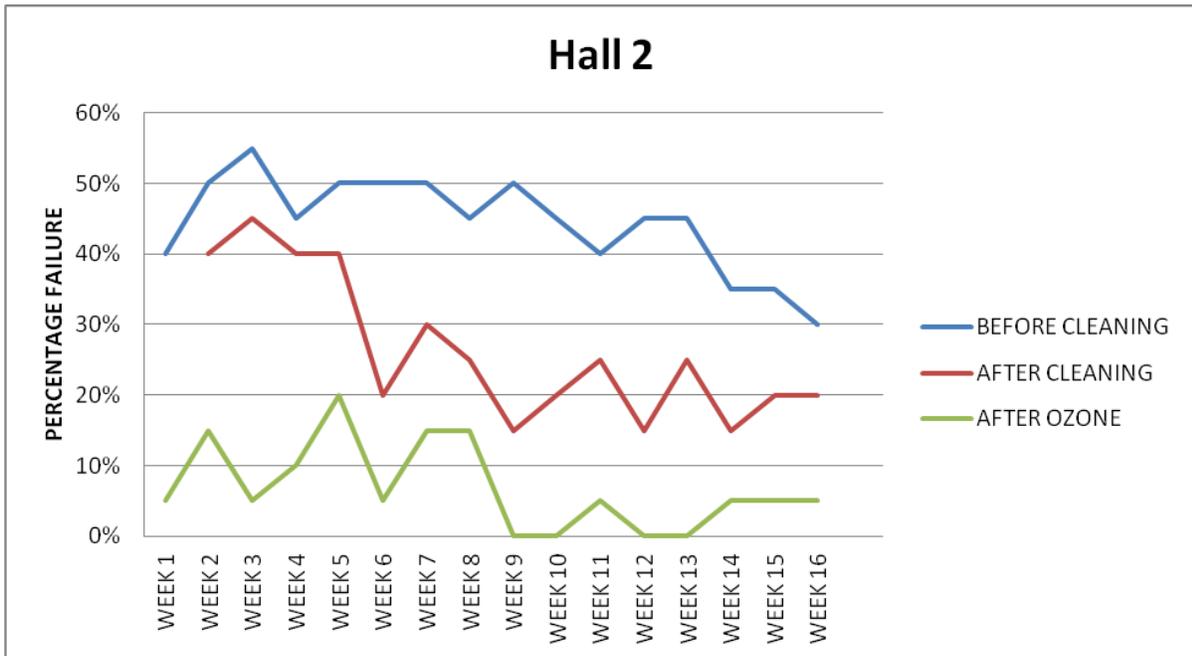
After week 16 there was an extended break in ozone treatment (week 17 – 35). This demonstrated that *Listeria* % positive sites returned to pre ozone treatment levels. However, Campden BRI has been informed that its re-instigation in one Hall “brought the *Listeria* spp positives down to levels seen prior to stopping production and ozonation, i.e. week 16”.

Campden BRI had no part in the field trial and the data has kindly been released to us for publication by the validating factory. The samples were taken by trained factory personnel before and after cleaning and after ozone application. The Petrifilm results were analysed by the company in question.

Graph 3.2.3a Hall 1: % Listeria spp. positives over 16 week period



Graph 3.2.3b Hall 2: % Listeria spp. positives over 16 week period



4. CONCLUSIONS

Gaseous ozone in a high humidity atmosphere has been shown to reduce the population levels of a range of environmental and pathogenic organisms, both in laboratory trials when dried on to surfaces and in field trials carried out in various production facilities (e.g. high care sandwich making or dry food production areas such as dough manufacturing).

There is a relationship between ozone concentration, contact time, type of micro-organisms present and log reduction achieved; however, in laboratory trials there seems to be no practical difference in the reduction achieved due to test surface location or orientation, or in restricted access exposures (w-tubes – laboratory trials).

The results suggested that, for each microorganism tested, it could be possible to describe a relationship between ozone concentration and exposure time that can be described as an ozone dosage.

As an overall conclusion from the laboratory trials ozone has several advantages; it can effectively penetrate every part of a room, including sites that might prove difficult to gain access to with conventional liquids and manual disinfection procedures. The major disadvantage of using gases, such as ozone, is the potential toxicity at high concentrations, which precludes using them in areas where people are working. The technique can therefore only be used in areas that can be isolated and sealed off during the decontamination process.

The number of treatments over time has been shown to be important for food contact and environmental surfaces. Single applications or applications over two consecutive days were shown to have a limited effectiveness (Malinowska and Holah, 2007). However, studies carried out over longer periods (≥ 3 applications) demonstrated a downward trend after applications over 3 consecutive days use (3 applications total).

The results for the pizza factory (3.2.1) after 3 days indicated a downward trend in the numbers of microorganisms present on food contact and environmental surfaces, both before cleaning and after cleaning and disinfection, throughout. This was supported by the 4 week trial at the sandwich factory (3.2.2) and further supported by two 16-week field trials in a dry foods dough manufacturing facility (3.2.3).

Ozone generation equipment manufacturers have postulated that when ozone is first applied to a cleaned room, there is a mass of organic material that creates an ozone demand which must be satisfied by oxidation before any significant oxidation of microorganisms can occur. In essence, this is no different from the effect of organic matter on traditional oxidising chemical disinfectants, e.g. a chlorine organic break point in water treatment.

During the 4 week (3.3.2: 8 ppm 30 minutes) and 16 week (3.3.3: 6 ppm 4 hours) trials no adverse effects were observed by ozone on the structure and fabric of the building. The management of the factory have also reported no adverse effects over the time the ozonation equipment has been installed.

The results of field trials demonstrate that, to be effective in a production environment (even one that has been cleaned), ozone requires at least 3 applications; however, once it starts to be effective successive cleaning and ozone use results in a continuous downward trend in counts which carries over, reducing detectable organisms in the environment prior to cleaning/disinfection (Graphs 3.2.3a & b). If the application is stopped levels of detectable pathogens can increase (Graph 3.2.3a).

Overall, therefore, the results of available laboratory data and field trial studies demonstrate that ozone has the potential to be an effective environmental disinfectant. However, any use of ozone as an addition to normal cleaning and disinfection practices or a replacement for chemical disinfection must be appropriately validated for each factory situation.

5. REFERENCES:

- Anon (2001) EN 13697:2001 Chemical disinfectants and antiseptics – Quantitative non-porous surface test for the evaluation of bactericidal and/or fungicidal activity of chemical disinfectants used in food, industrial, domestic and institutional areas – Test methods and requirements without mechanical action (phase 2/step 2) <http://www.bsigroup.com/>
- Bermudez-Aguirre, D. & Barbosa-Canovas, G.V. (2013) Disinfection of selected vegetables under non-thermal treatments: chlorine, citric acid, ultraviolet light and ozone, *Food Control* 29 (1), 2013, 82-90
- Brandit, J. (2009) The case for ozone, *Food Quality* (Dec/Jan)
- Boisrobert, C. (2002) US regulatory review of the use of ozone in the food industry, *Agricultural and Food Processing Applications of ozone as an antimicrobial*
- Dusseau, J.Y., Duroselle, P. & Freney, J. (2012) Gaseous Sterilization (Chapter 15.3) *Principles and Practice of Disinfection, Preservation and Sterilization* 5th Edition ed Fraise, Maillard, Sattar, Wiley-Blackwell 316-328
- Fan, L., Song, J., McRae, K.B., Walker, B.A. & Sharpe, D. (2007) Gaseous ozone treatment inactivates *Listeria innocua in vitro*. *Journal of Applied Microbiology* 103, 2657-2663.
- Health & Safety Executive (1983) Ozone: health hazards and precautionary measures. HSE Guidance Note - EH 38.
- Health & Safety Executive (2005) Table 1: List of approved workplace exposure limits. HSE - EH 40.
- Holah, J., Bird, J. & Hall, K. (2002) The microbial ecology of high risk, chilled food factories; evidence for persistent *Listeria* spp. and *Escherichia coli* strains. R&D Report No.165, Campden BRI.
- Holah, J., Bird, J. & Hall, K. (2004) *Listeria monocytogenes* and *Escherichia coli* in high risk, chilled food factories; where do they come from? R&D Report No.199, Campden BRI.
- Hudson, J.B., Sharma, M. & Petric, M. (2007) Inactivation of norovirus by ozone gas in conditions relevant to healthcare. *Journal of Hospital Infection* 66, 40-45.
- Kim, J.-G. & Yousef, A.E. (2000) Inactivation kinetics of foodborne spoilage and pathogenic bacteria by ozone. *Journal of Food Science* 65(3), 521-528.

- Maillard, J-Y. Sattar S.A. Pinto, F. (2012). Virucidal activity of microbicides. In: Eds Fraise, A. P. Maillard, J-Y. Sattar, S.A. *Russell, Hugo & Ayliffes Principles and Practice of Disinfection, Preservation and Sterilization*. Wiley-Blackwell pp 195
- Malinowska, A. and Holah, J.T. (2007) Whole room disinfection - potential for environmental pathogen control, *New Food* (no5) 22-26
- Middleton, K. (2009) Whole room disinfection. *Food and Beverage International* 8(6), 46-47
- Middleton, K. (2010) Whole room disinfection : efficacy of whole room disinfection methods including ozone. R&D report 299, Campden BRI
- Moore, G., Griffiths, C. & Peters, A. (2000) Bactericidal properties of ozone and its potential application as a terminal disinfectant. *Journal of Food Protection* 63(8),1100-1106
- Pascaul, A., Llorca, I. & Canut, A. (2007) Use of ozone in food industries for reducing the environmental impact of cleaning and disinfection activities, *Trends in Food Science & Technology* 18 (Suppl. 1) s29-s35
- Selma, M. V., Ibanez, A.M., Cantwell, M. & Suslow, T. (2008) Reduction by gaseous ozone of *Salmonella* and microbial flora associated with fresh-cut cantaloupe. *Food Microbiology* 25(4) 558-565
- Shah N.N.A.K. Rahman, R.A. Chuan, L.T. (2011), Application of gaseous ozone to inactivate *Bacillus cereus* in processed rice. *Journal of Food Process Engineering* 34(6), 2220-2232
- Siqueria Lanita, C. de. & Botelho da Silva, S. (2008) Use of ozone in industrial cold rooms to control yeast and moulds during parmesan cheese ripening. *Brazilian Journal of Food Technology* 11(3), 182
- Taylor, J. & Chana, D. (2000) The evaluation of ozone for airborne and surface disinfection. R&D Report No.109, Campden BRI.
- Troller Pinto, A. Scmidit, V. Aparecida Raimundo. S. (2013) Mould control by ozonation in ripening cheese room. *Acta Scientiae Veterinariae* 35 (3), 333-337
- Xuetong Fan, Sokoral, K. J. B., Engermann, J., Gutler, J.B. & Yanhong Liyu (2012) Inactivation of *Listeria*, *Salmonella* Typhimurium and *Escherichia coli* O157:H7 on surface and stem scar areas of tomatoes using in package ozonation. *Journal of Food Protection* 75(9), 1611-1618

Zhang Chunyan, Cai Jingping & Pan Feng. (2007) Study of microbial contamination condition of low-moisture capsicum powder and control technology. Food Science China 28(1), 131-134